



## FBC WORKSHEET 2010

The samples have been tested and found to be homogeneous and stable for the purpose of these exercises.  
These samples are supplied in liquid form – no reconstitution is required.

<b>JANUARY</b>	Cycle 15 Run 1	FB15-01a FB15-01b	WCC, RCC, Hb, Hct, MCV, Plt WCC, RCC, Hb, Hct, MCV, Plt
<b>FEBRUARY</b>	Cycle 15 Run 2	FB15-02a FB15-02b	WCC, RCC, Hb, Hct, MCV, Plt WCC, RCC, Hb, Hct, MCV, Plt
<b>MARCH</b>	Cycle 15 Run 3	FB15-03a FB15-03b	WCC, RCC, Hb, Hct, MCV, Plt WCC, RCC, Hb, Hct, MCV, Plt
<b>APRIL</b>	Cycle 15 Run 4	FB15-04a FB15-04b	WCC, RCC, Hb, Hct, MCV, Plt WCC, RCC, Hb, Hct, MCV, Plt
<b>MAY</b>	Cycle 15 Run 5	FB15-05a FB15-05b	WCC, RCC, Hb, Hct, MCV, Plt WCC, RCC, Hb, Hct, MCV, Plt
<b>JUNE</b>	Cycle 15 Run 6	FB15-06a FB15-06b	WCC, RCC, Hb, Hct, MCV, Plt WCC, RCC, Hb, Hct, MCV, Plt
<b>JULY</b>	Cycle 16 Run 1	FB16-07a FB16-07b	WCC, RCC, Hb, Hct, MCV, Plt WCC, RCC, Hb, Hct, MCV, Plt
<b>AUGUST</b>	Cycle 16 Run 2	FB16-08a FB16-08b	WCC, RCC, Hb, Hct, MCV, Plt WCC, RCC, Hb, Hct, MCV, Plt
<b>SEPTEMBER</b>	Cycle 16 Run 3	FB16-09a FB16-09b	WCC, RCC, Hb, Hct, MCV, Plt WCC, RCC, Hb, Hct, MCV, Plt
<b>OCTOBER</b>	Cycle 16 Run 4	FB16-10a FB16-10b	WCC, RCC, Hb, Hct, MCV, Plt WCC, RCC, Hb, Hct, MCV, Plt
<b>NOVEMBER</b>	Cycle 16 Run 5	FB16-11a FB16-11b	WCC, RCC, Hb, Hct, MCV, Plt WCC, RCC, Hb, Hct, MCV, Plt
<b>DECEMBER</b>	Cycle 16 Run 6	FB16-12a FB16-12b	WCC, RCC, Hb, Hct, MCV, Plt WCC, RCC, Hb, Hct, MCV, Plt

## **SAMPLE HANDLING AND PROCESSING**

**These instructions were obtained directly from the manufacturer of the control material.**

The samples provided contain 1mL of stabilised whole blood. Store the vials upright at 2-8°C when not in use. **Protect vials from overheating and freezing.** Unopened vials are stable until the expiration date.

1. Remove vial from the refrigerator and allow to warm at room temperature for 15 minutes before mixing.
2. To mix, hold vial horizontally between palms of the hands. **Do not pre-mix on a mechanical mixer.**
  - Roll the vial back and forth for 20-30 seconds; occasionally invert the vial. Mix vigorously but do not shake.
  - Continue to mix in this manner until the red cells are completely suspended. Vials stored for a long time may need extra mixing.
  - Gently invert the vial 8-10 times immediately before running each sample.
3. Return vials to refrigerator within 30 minutes of use.

## **PARTICIPANTS WITH MULTIPLE INSTRUMENTS**

- If your laboratory has more than one backup instrument, the sample may be insufficient to process through the primary mode. In cases like this, please process your samples through the secondary mode.
- It is the participant's responsibility to be consistent in the submission of results. The instrument designated xxxx.1 should remain the same throughout the cycle in order for the end of cycle statistics and cumulated data to be meaningful.

## **BAYER USERS**

- Laboratories processing their QAP-FBC samples on Bayer Instrumentation should run a saline primer after aspirating the RCPA Haematology QAP-FBC samples. This is done to prevent carryover of the platelets, which has occurred in the past and is unique to the product and the Bayer instrumentation.

## **SYSMEX / CELL DYN USERS**

- As a result of the Primary/Secondary Mode special exercise sent in 2004 we recommend all Sysmex users process the QAP samples through the OPEN/MANUAL MODE.
- Sysmex XE/XT: The reticulocyte channel should NOT be enabled.
- Sysmex XE/XT: Please report your Impedance count for all parameters.

## **CELL DYN USERS**

- CD4000: Please report your "Optical" count for the Platelets.